

## Toxic and repellent effects of three botanical oils against adult *Aphis craccivora* Koch. (Homoptera: Aphididae) under laboratory conditions

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**Abstract:** In the present study, three botanical oils viz. neem, karanja and mahogany were tested against the adult bean aphid, *Aphis craccivora* Koch to evaluate the toxic and repellent effects under laboratory condition in the Department of Entomology, Hajee Mohammad Danesh Science and Technology University, Dinajpur during September 2016 to January 2017. The results indicated that all the botanical oils had different levels of toxic and repellent effects and was found effective for controlling the aphid. Among the botanicals, the highest (29.83%) mortality was found in mahogany oil whereas the lowest (22.96%) in karanja oil against the adult aphid. Mahogany oil also showed the highest (77.33%) repellent effects i.e. class IV among the three tested botanical oils. On the contrary, karanja oil showed the lowest repellent (34.00%) effects, i.e. class II and neem oil belongs to the repellency class III. However, with the finding of this study, it may conclude that for eco-friendly management of *A. craccivora*, all the botanical oils showed promising toxic and repellent effects against the adult bean aphid but mahogany oil was the highest. So, mahogany oils may be considered at farmer's level as it is cheaper, easily available, processable, and usable for the management of bean aphid.

**Key words:** *Aphis craccivora*, botanical oils, mortality, repellent effect.

### Introduction

Country bean (*Lablab purpureus* L.) locally known as deshi sheem is a highly proteinaceous legume and a major winter vegetable in Bangladesh (Salim *et al.*, 2013). The amounts of crude protein of the country bean were estimated 21 to 38% and 20 to 28% in leaf and seed, respectively (Cook *et al.*, 2005). Besides, seeds have some micronutrients and minerals also (Kala *et al.*, 2010; Shaahu *et al.*, 2015). It is the atmospheric N-fixing legume and also a valuable green manure forming crop (Adebisi & Bosch, 2004). However, crops are threatened by several insect pests resulting huge economic losses (Thejaswi *et al.*, 2008; Oliveira *et al.*, 2014).

Among the insect pests of legumes crops, bean aphid, *Aphis craccivora* Koch (Homoptera: Aphididae) is most destructive and cosmopolitan pests (Capinera, 2001; Madahi *et al.*, 2013). There are about 4700 species of aphids in the world which cause enormous loss of vegetables (Blackman & Eastop, 2007; Alikhani *et al.*, 2010). Both the adults and nymphs suck plant sap and cause up to 10-90% yield loss (Akhtar *et al.*, 2010; Razaq *et al.*, 2011). They damage all parts of the plants except roots and show symptoms as stunting growth, crinkling and curling of leaves, pods and indirectly by transmission of plant viruses (Saranya *et al.*, 2010; Sainsbury *et al.*, 2010; Kamphuis *et al.*, 2012). They also

secrete honeydew causing the growth of sooty mould fungus, which inhibits photosynthesis (Kingler *et al.*, 2001; Smith & Boyko, 2007; Singh *et al.*, 2014).

Aphid is a prolific breeder and randomly corrosive insecticides are used to control them for quick knockdown (Pavela *et al.*, 2009). But chemical protection measures exert so many serious drawbacks (Karungi *et al.*, 2000; Lee *et al.*, 2001; Ambethar, 2009). Their extensive and indiscriminate use causes ecological imbalance, resistance to pest, pest resurgence, outbreaks of secondary pests and also creates phytotoxicity, residues in foods and feeds, killing beneficial organisms in the ecosystem (Mahmud *et al.*, 2002; Ashamo, 2004; Stapel *et al.*, 2000). Hence, researchers and scientists all over the world are now trying to adopt alternatives of insecticides to protect crop from insect pest (Rajappan *et al.*, 2000; Isman, 2006). Use of natural products are an excellent source for controlling insect pests (Gorur *et al.*, 2008; Franck *et al.*, 2009). Botanicals become promising to control pests by offering several advantages in comparing to insecticides (Attia *et al.*, 2013; Kedia *et al.*, 2015). Essential plant oils demonstrated high insecticidal activity against insect pests (Cheng *et al.*, 2003; Ahmedani *et al.*, 2007; Khater, 2012). Therefore, the present study was undertaken to evaluate the effectiveness of three botanical oils viz. neem, karanja and mahogany for controlling bean aphids.

## Materials and Methods

### Experimental site

The experiment was carried out in the laboratory conditions ( $25 \pm 5^\circ\text{C}$ , 65-75% RH) of the Department of Entomology, Hajee Mohammad Danesh Science and Technology University (HSTU), Dinajpur, Bangladesh during the period from September 2016 to January 2017.

### Raising of host plant

Country bean used as host plant were raised in small plastic pots (11 cm x 14 cm) in the corridor of the laboratory. Plants were fertilized as needed and watering was done regularly. No pesticide was applied during the experiment.

### Collection and rearing of test insects

*A. craccivora* were collected from the infested bean plants of HSTU central farm. Twigs of bean harboring with *A. craccivora* colonies were brought to the laboratory ( $25 \pm 5^\circ\text{C}$ ,  $70 \pm 5\%$  RH). Afterwards, aphids were gently removed from the bean twigs with the help of soft camel hair brush and released on the fresh bean twigs kept in rectangular jars (14 cm x 10.5 cm x 30 cm) in the laboratory. The jars were covered with a piece of cloth and fastened with rubber bands to prevent insect's getaway. Newly hatched crawlers were collected from the laboratory culture and were placed on to the fresh bean twigs for mass culture. These processes were continued upto the experimental requirements.

### Identification of adult *A. craccivora*

Traditionally, aphid species have been identified based on their morphological characters (Emden & Harrington, 2007). Adults were identified by using the procedures as stated by Blackman & Eastop (2000).

### Tested botanical oils

Commercially available three plant oils namely neem (*Azadirachta indica*), mahogany (*Swietenia mahagoni* L.) and karanja oil (*Pongamia pinnata* L.) were collected from the local market of Dinajpur town. Four concentrations (0.5, 1.0, 1.5, and 2.0%) were prepared with distilled water containing 0.1% detergent as emulsifier.

### Direct toxicity test

Toxicity and repellent effects were performed in the ambient laboratory conditions against adult *A. craccivora*. Fresh young bean leaves were collected and dipped in assigned tested oils for 5-10 seconds containing four

concentrations (0.5, 1.0, 1.5, and 2.0%) and then the leaves were air-cured for 15 m. Each leaf separately placed in each Petri dish (90 mm). Ten (10) aphids (both adults and nymphs separately) were released on each leaf twig by the help of a camel-hair brush. Three replications were done for each concentration. Adult and nymph mortalities were recorded at 24, 48 and 72 hour after treatment (HAT). Mortality data were corrected by Abbott's (1925) formula:

$$P = \frac{P' - C}{100 - C} \times 100$$

Where,  $P$  = Percentage of corrected mortality

$P'$  = Observed mortality (%)

$C$  = Mortality (%) at control.

### Repellency test

The repellency test was conducted according to the method as described by Talukder & Howse (1995). Petri dishes (120 mm) sized filter papers (Whatman No. 40) were cut in two half and 1.0 ml of each concentrate oils were applied to a half filter paper uniformly with a pipette. The treated half were then air-dried and attached with the untreated half with a cello-tape those could not interfere with the free movement of insect from one half to another. Distance between the filter paper segments remained sufficient to prevent seepage of test samples from one half of circle to another. Each filter paper was then placed in a Petri-dish and 5 pair adults and nymphs were released at the center of the filter paper. Three replications were done for each concentration for each plant extract. Number of insects on each treated and untreated portion was counted at two hour intervals up to the 10<sup>th</sup> hour.

The data were expressed as percentage repulsion (PR) by the following formula:  $[\text{PR} (\%) = (\text{Nc}-50) \times 2]$ . Where,  $\text{Nc}$  = the percentage of insects present in the control half. Positive (+) values expressed repellency while negative (-) attractancy. The average values were then categorized according to the following scale (McDonald *et al.* 1970).

Class	Repellency (%)	Class	Repellency (%)
0	>0.01-0.1	III	40.1-60
I	0.1-20	IV	60.1-80
II	20.1-40	V	80.1-100

### Statistical Analysis

The data were analyzed using completely randomized design (CRD) through Mathematical and Statistical program (MSTAT-C). All graphs were done using MS-Excel software. The treatment mean values were compared by Duncan's New Multiple Range Test (DMRT). The median lethal values (LD<sub>50</sub>) were determined by probit analysis.

### Results

#### Direct toxic effect of oils on adult *A. craccivora*

The toxicity effects of three botanical oils, doses and interactions against the adult of *A. craccivora* are shown in the Tables 1-3. The highest (29.83%) adult mortality was recorded in mahogany oil while the lowest (22.96%) in karanja oil (Table 1). From the result it was

observed that the order of toxicity of three oils were mahogany > neem > karanja. The highest mortality (51.63%) was found at the dose 2.0% (Table 2). Conversely, the lowest mortality (17.72%) was observed at the dose 0.5%. Average mortality was increased with increased dose and time. In the interaction of oils, doses and times, significantly the highest mortality (62.70%) was observed at 72 HAT in mahogany oil at 2.0% while the lowest at 0.0 (control) (2.23%) (Table 3). The LD<sub>50</sub> values of the tested oils against *A. craccivora* adult showed that mehogany oil possessed the lowest LD<sub>50</sub> values at 24, 48 and 72 HATs (Table 4). Conversely, karanja oil showed the highest LD<sub>50</sub> values at 24, 48 and 72 HATS. It was observed that the LD<sub>50</sub> values for mehogany oil were 2.75%, 1.73%, 1.04% at 24, 48 and 72 HATs respectively, among the tested botanicals (Table 4). The probit mortality data and log doses of the tested oils were presented in Figures 1.

**Table 1:** Direct toxic effect of different botanical oils against the adult of *A. craccivora* at different HAT\* (oils × times)

Treatments (Oils)	Adult mortality (%) at different HAT			Average mortality (%)
	24	48	72	
Neem	16.00 <sup>ab</sup>	21.85 <sup>b</sup>	34.37 <sup>b</sup>	24.07 <sup>b</sup>
Karanja	14.00 <sup>b</sup>	23.26 <sup>ab</sup>	31.63 <sup>b</sup>	22.96 <sup>b</sup>
Mahogany	19.33 <sup>a</sup>	27.48 <sup>a</sup>	42.67 <sup>a</sup>	29.83 <sup>a</sup>
LSD	4.01	5.187	4.480	3.750
CV %	32.68	28.74	16.58	19.63

\*HAT= Hours after treatment. Within column values followed by different letter(s) are significantly different by DMRT at 5% level of probability.

**Table 2:** Toxicity effect of different doses of botanical oils against the adult of *A. craccivora* at different HAT\* (oils × times)

Treatments (Doses %)	Adult mortality (%) at different HAT			Average mortality (%)
	24	48	72	
2.0	34.44 <sup>a</sup>	51.48 <sup>a</sup>	69.01 <sup>a</sup>	51.63 <sup>a</sup>
1.5	21.11 <sup>b</sup>	33.09 <sup>b</sup>	50.49 <sup>b</sup>	34.90 <sup>b</sup>
1.0	13.33 <sup>c</sup>	18.27 <sup>c</sup>	33.21 <sup>c</sup>	21.60 <sup>c</sup>
0.5	13.33 <sup>c</sup>	14.81 <sup>c</sup>	25.06 <sup>d</sup>	17.72 <sup>c</sup>
Control	0.00 <sup>d</sup>	3.33 <sup>d</sup>	3.33 <sup>e</sup>	2.23 <sup>d</sup>
LSD	5.175	6.696	5.783	4.841
CV %	32.68	28.74	16.58	19.63

\*HAT= Hours after treatment. Within column values followed by different letter(s) are significantly different by DMRT at 5% level of probability.

**Table 3:** Direct toxic effect of botanical oils at different doses against the adult of *Aphis craccivora* at different HAT (oils x doses x times)

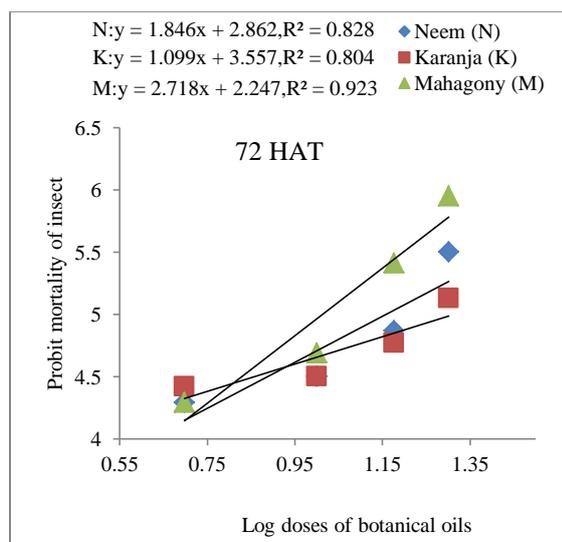
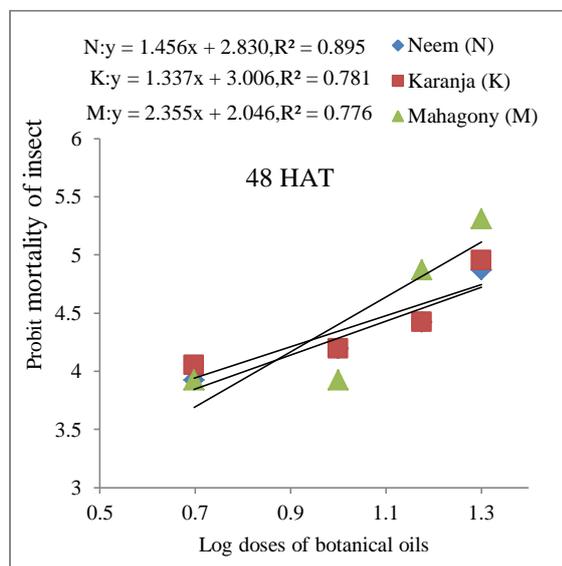
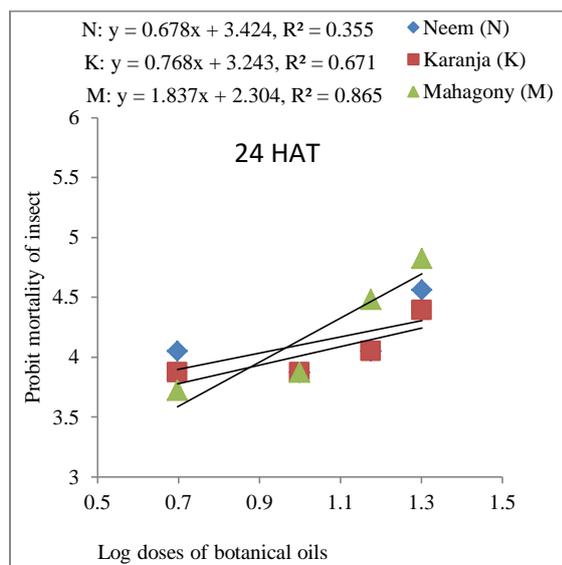
Treatments (Oils)	Doses (%)	Adult mortality (%) at different HAT*			Average mortality (%)
		24	48	72	
Neem	2.0	33.33 <sup>b</sup>	44.44 <sup>b</sup>	68.89 <sup>b</sup>	48.87 <sup>b</sup>
	1.5	16.67 <sup>c</sup>	27.41 <sup>c</sup>	44.81 <sup>d</sup>	29.63 <sup>c</sup>
	1.0	13.33 <sup>c</sup>	20.37 <sup>cd</sup>	30.74 <sup>efg</sup>	21.47 <sup>cde</sup>
	0.5	16.67 <sup>c</sup>	13.70 <sup>de</sup>	24.07 <sup>g</sup>	18.13 <sup>e</sup>
Karanja	2.0	26.67 <sup>b</sup>	48.15 <sup>b</sup>	55.19 <sup>c</sup>	43.33 <sup>b</sup>
	1.5	16.67 <sup>c</sup>	27.04 <sup>c</sup>	41.11 <sup>de</sup>	28.27 <sup>cd</sup>
	1.0	13.33 <sup>c</sup>	20.74 <sup>cd</sup>	31.11 <sup>efg</sup>	21.73 <sup>cde</sup>
	0.5	13.33 <sup>c</sup>	17.04 <sup>cd</sup>	27.41 <sup>fg</sup>	19.23 <sup>de</sup>
Mahogany	2.0	43.33 <sup>a</sup>	61.85 <sup>a</sup>	82.96 <sup>a</sup>	62.70 <sup>a</sup>
	1.5	30.00 <sup>b</sup>	44.81 <sup>b</sup>	65.56 <sup>b</sup>	46.80 <sup>b</sup>
	1.0	13.33 <sup>c</sup>	13.70 <sup>de</sup>	37.78 <sup>def</sup>	21.60 <sup>cde</sup>
	0.5	10.00 <sup>c</sup>	13.70 <sup>de</sup>	23.70 <sup>g</sup>	15.80 <sup>e</sup>
	0.0(Control)	0.00 <sup>d</sup>	3.33 <sup>e</sup>	3.33 <sup>h</sup>	2.23 <sup>f</sup>
LSD		8.963	11.60	10.02	8.385
CV (%)		32.68	28.74	16.58	19.63

\*HAT= Hours after treatment, within column values followed by different letter(s) are significantly different by DMRT at 5% level of probability.

**Table 4:** Relative toxicity (probit analysis) of different oils against *A. craccivora* adults after 24, 48 and 72 hours

Botanical oils	Adult used (n)	LD <sub>50</sub> (%) values	95% fiducial limits		$\chi^2$ values (df=2)
			Lower	Upper	
24 HAT*					
Neem	30	14.72	0.2425	893.84	2.57
Karanja	30	17.23	0.2347	1266.10	0.81
Mahogany	30	2.75	1.5167	4.98	1.25
48 HAT*					
Neem	30	3.01	1.3544	6.6945	0.93
Karanja	30	2.99	1.2690	7.0778	1.77
Mehogany	30	1.73	1.3161	2.2943	4.19
72 HAT*					
Neem	30	1.42	1.0481	1.9342	2.80
Karanja	30	2.04	0.9881	4.2299	1.00
Mehogany	30	1.04	0.8481	1.2872	2.20

\*HAT = Hour after treatment.



**Figure 1.** Relationship between probit mortality and log doses of different botanical oils on *A. craccivora* adult at various exposure times.

**Repellent effect of oils against *A. craccivora* adults**

The repellent effect of three botanical oils on the adults of bean aphid are presented in Table 5. Among the three tested oils, mahagony and karanja showed the highest (77.33%) and the lowest (34.0%) repellency, respectively. Repellency percent decreased with the progress of time and increased with increasing doses of all oils. The repellency class of different oils at different concentration levels varied between II to V. Among the botanicals the highest repellency effect was found at 2.0 % mehogany oil (84.0%) whereas the lowest (26.67 %) at doses (1.0 and 1.5%) of karanja oil (Table 5).

**Discussion**

Bean aphid, *A. craccivora* have become schedule pests of bean and causes major threat to its production in Bangladesh. Insecticides are solely relying to reduce its economic losses during production as a part of integrated management strategies against this notorious pest. However, the accumulation of toxic compounds in the green pods makes it vulnerable for human consumption as a delicious vegetable. Synthetic insecticides have been found effective in managing bean aphid to minimize yield losses but so many serious drawbacks. Results of the present study revealed that all the tested oils had toxic and repellent effects against the adult, *A. craccivora* under laboratory conditions. Among the botanicals, the highest (29.83%) mortality was found in mahagony oil whereas the lowest (22.96%) in karanja oil against the adults bean aphid. The mahagony oil offered promising toxicity by applying the highest dose of 2.0% against adult aphids. The results of the present findings are also in close proximity with those of Pinto *et al.* (2013). They concluded that botanical oils of neem (Neemseto ®), and cotton seed being achieved satisfactory control while 100% on the thiamethoxam-treated plants infested by *Aphis gossypii* Glover. The present results are similar with Patil & Chavan (2009). They cited that *Acacia concianna* extracts were the most toxic against sugarcane woolly aphid, *Ceratozacuna lanigera* Zehnter. Mortality of the tested extracts was concentrations and exposure time dependent under laboratory conditions. Lin *et al.* (2009) reported that sugar apple, (*Annona squamosa*) seed oil, an edible tropical fruit was also very effective in controlling the cotton aphid, *A. gossypii* Glover on melon leaves.

**Table 5:** Repellent effect of different botanical oils and doses against the adult *A. craccivora* at different HAT\*

Treatments Doses (Oils)	Repellency (%) at different HAT*	Repellency (%) at different HAT*					Average	Class
		(%)	2	4	6	8		
Neem	2.0	60.00 <sup>bc</sup>	66.67 <sup>ab</sup>	66.67 <sup>ab</sup>	73.33 <sup>ab</sup>	73.33 <sup>ab</sup>	68.00 <sup>ab</sup>	IV
	1.5	86.67 <sup>ab</sup>	73.33 <sup>ab</sup>	40.00 <sup>a-c</sup>	66.67 <sup>a-c</sup>	53.33 <sup>bc</sup>	64.00 <sup>ab</sup>	IV
	1.0	73.33 <sup>a-c</sup>	73.33 <sup>ab</sup>	40.00 <sup>a-c</sup>	40.00 <sup>b-d</sup>	53.33 <sup>bc</sup>	56.00 <sup>bc</sup>	III
	0.5	46.67 <sup>cd</sup>	26.67 <sup>c</sup>	33.33 <sup>bc</sup>	23.33 <sup>d</sup>	26.67 <sup>cd</sup>	31.33 <sup>d</sup>	II
Karanja	2.0	33.33 <sup>d</sup>	46.67 <sup>bc</sup>	46.67 <sup>a-c</sup>	46.67 <sup>a-d</sup>	26.67 <sup>cd</sup>	40.00 <sup>cd</sup>	II
	1.5	26.67 <sup>d</sup>	46.67 <sup>bc</sup>	46.67 <sup>a-c</sup>	33.33 <sup>cd</sup>	33.33 <sup>cd</sup>	37.33 <sup>cd</sup>	II
	1.0	26.67 <sup>d</sup>	26.67 <sup>c</sup>	33.33 <sup>bc</sup>	33.33 <sup>cd</sup>	40.00 <sup>cd</sup>	32.00 <sup>d</sup>	II
	0.5	33.33 <sup>d</sup>	26.67 <sup>c</sup>	26.67 <sup>c</sup>	26.67 <sup>d</sup>	20.00 <sup>d</sup>	26.67 <sup>d</sup>	II
Mehogony	2.0	93.33 <sup>a</sup>	86.67 <sup>a</sup>	73.33 <sup>a</sup>	80.00 <sup>a</sup>	86.67 <sup>a</sup>	84.00 <sup>a</sup>	V
	1.5	73.33 <sup>a-c</sup>	86.67 <sup>a</sup>	73.33 <sup>a</sup>	73.33 <sup>ab</sup>	73.33 <sup>ab</sup>	76.00 <sup>ab</sup>	IV
	1.0	86.67 <sup>ab</sup>	73.33 <sup>ab</sup>	73.33 <sup>a</sup>	66.67 <sup>a-c</sup>	73.33 <sup>ab</sup>	74.67 <sup>ab</sup>	IV
	0.5	66.67 <sup>a-c</sup>	80.00 <sup>ab</sup>	66.67 <sup>ab</sup>	73.33 <sup>ab</sup>	86.67 <sup>a</sup>	74.67 <sup>ab</sup>	IV
LSD		25.12	31.78	34.17	30.89	26.35	20.72	
CV		25.31	31.72	39.24	34.55	29.01	22.20	

\*HAT = Hour after treatment. Mean followed by different letter(s) are significantly different by DMRT at 5% level of probability

Significant level of success in the suppression of aphid reported by various authors with extracts of different botanicals including neem and karanja leaf (Saikia *et al.*, 2000; Katsvangwa & Chigwaza, 2004; Rawleigh & Boyd, 2008; Biswas, 2013). The result of this study indicates that all levels of tested oil except 0.5% and 1.0% had shown sufficient aphicidal effect.

More current research illustrated that various plant products have been tried by several researchers with a good degree of success as protectants against several species of aphids. The results of the present study are comparable with various scholars where they mentioned that different extracts of Mexican marigold, sodom apples, garlic and ginger were promising against the aphids *Brevicoryne brassicaea* (Peris & Kiptoo, 2017); neem and moringa (*Moringa olifera*) to wheat aphid *Schizaphis graminum* (R.), *Rhopalosiphum padi* (L.) and *Sitobion avenae* (F.) (Shah *et al.*, 2017); Orange peel (*Citrus sinensis*), Bitter goard (*Momordica dioica*), garlic (*Allium vineale*), mari gold; hot pepper (*Capsicum frutescens*) and tobacco (*Nicotiana tabacum*) to wheat aphid (Iqbal, 2011); garlic bulbs (*Allium sativum*), endod (*Phytolacca dodecandra*) and neem seeds (*Azadirachta*

*indica*) to pea aphids, *Acyrtosiphon pisum* (Harris) (Megersa, 2016).

Among all the tested botanicals, mehogony, neem and karanja attributed significant repellency against *A. craccivora*. Varied repellency of different tested botanical oils indicated that the pest suppressing properties are not uniformly distributed. Ginger extract has been used to control aphids in various infested plants of Brassica family. It has repellent mode of action against the aphids but no effect on non-target organisms e.g. parasitoids. Suthisut *et al.* (2011) experienced that *A. conchigera* and *C. zedoaria* extracted essential oils at the highest concentration ( $314.56 \times 10^{-3} \mu\text{l}/\text{cm}^2$ ) repelled *S. zeamais* adults, with > 87% of the insects found on the untreated filter paper. Besides, aphid populations suppressed more effectively when it was applied in combined with neem and eucalyptus, *Eucalyptus globulus* (family Myrtaceae) leaves extract (Manzoor *et al.*, 2015; Ali *et al.*, 2015).

The biological activities of tested oils can be ascribed to several alkaloid contents as an insecticidal potency (Jastad *et al.*, 2009; Alice *et al.*, 2007). The alkaloids, terpenoids, steroids, glycosides as Cyclomahogenol, tannins, alkaloids, saponins and terpenoids as

main phytoconstituents present in mehogany (Chakraborty & Basak, 1971; Sahgal *et al.*, 2009; Hajra *et al.*, 2011). Karanja oil contains karanjin another alkaloid (Prakash & Rao, 1996) having insect control properties against pulse beetle, *Callosobruchus chinensis* (Hossain *et al.*, 2014). These chemical compounds might associate with deterrent, repellent and anti-feeding actions against *A. craccivora*, *B. brassicaea* (Homoptera: Ahididae) but seems harmless or tolerated by beneficial natural enemies as well (Katsvangwa & Chigwaza, 2004). Spray of botanicals particularly neem seed oil and neem seed extracts did not reduce the parasitism rate of *Diaeretiella rapae* (McIntosh) (Hymenoptera: Braconidae) on green peach aphid, *M. persicae* under laboratory and field conditions (Lowery & Isman, 1995). In fact, plant based formulation especially neem not only shows systemic and contact actions but also acts as repellent, antifeedent, sterilents, growth inhibitors and toxicological repellents against insect pests, which has been considered as a low-cost management tactic (Gahukar, 2000). Some oils have broad spectrum insecticidal activity against aphid, affecting insect nervous and defence systems (Hold *et al.*, 2000; Isman, 2000; Ketoh, 2004). The biological activity of oils interferes with normal activities of insects resulting might to death (Schoonhoven, 1978).

### Conclusion

It is concluded that the botanical oils used in the present study had direct toxic effect on the adult and nymphs of *A. craccivora* which can be used as safe pesticides. Uses of these botanical oils have a great economic and environmental importance than the chemical pesticides. For this reason farmers can use botanical oils as insecticides in agricultural sector because of direct toxic and repellent effects against *A. craccivora* as well as other vegetable pests also. Among the three botanical oils mahogany showed the highest toxic and repellent effects against *Aphis craccivora*. The findings of the present investigation reveal the broad spectrum toxic properties of the tested indigenous botanical oils against the adult and nymph of *A. craccivora*. However, further studies need to be conducted to isolate and evaluate the compound with its mode of action of these oils.

### References

- Abbott, W.S. 1925. A method of computing the effectiveness of an insecticide. *Journal of the American Mosquito Control Association*. **3**: 302-303.
- Adebisi, A.A. & Bosch, C.H. 2004. *Lablab purpureus* (L.). Record from PROTA4U, Grubben, G.J.H., Denton, O.A. (Editors). PROTA (Plant Resources of Tropical Africa / Ressources végétales de l'Afrique tropicale), Wageningen, Netherlands.
- Ahmedani, M.S., Abdulkhalig, M.T., Tarig, M., Anwar, M. & Naz, S. 2007. Khapra beetle (*Trogoderma granarium* Everts) - A serious threat of food security and safety. *Pakistan J. Agril. Sci.* **44(3)**: 481-493.
- Akhtar, N., Ashfaque, M., Waseem, Gillani, W. A., Ata-ul-Mohsin, Tashfeen, A. & Begum, I. 2010. Antibiosis resistance in national uniform wheat yield trials against *Rhopalosiphum padi* (L.). *Pakistan J. Agril. Res.* **23(1-2)**: 59-63.
- Ali, H., Qasim, M., Saqib, H.S.A., Arif, M. & Islam S.U. 2015. Synergetic effects of various plant extracts as bio-pesticide against wheat aphid (*Diuraphis noxia* L.) (Hemiptera: Aphididae). *Afr. J. Agric. Sci. Technol.* **3**: 310-315.
- Alice, J., Sujeetha, R.P., Mathurani, N. & Nadarajan, L. 2007. Effect of indigenous plant products and oils against the pulse beetle, *Callosobruchus chinensis* (L.) on stored blackgram. *Asian J. Biol. Sci.* **2(1&2)**: 203-204.
- Alikhani, M., Rezwani, A., Rakhshani, E. & Madani, S.M.J. 2010. Survey of aphids (Hemiptera: Aphidoidea) and their host plants in central parts of Iran. *J. Entomol. Res.* **2(2)**: 7-16.
- Ambethar, V. 2009. Potential of entomopathogenic fungi in insecticide resistance management (IRM): a review. *J. Biopesticides.* **2(2)**: 177-193.
- Ashamo, M.O. 2004. Effects of some plant powders on the yam moth, *Dasyses rugosella stainton* (Lepidoptera: Tineidae). *Biosci. Biotech. Res. Commun.* **16**: 41-46.
- Attia, S., Grissa, K.L., Lognay, G., Bitume, E., Hance, T. & Mailleux, A.C. 2013. A

- review of the major biological approaches to control the worldwide pest *Tetranychus urticae* (Acari: Tetranychidae) with special reference to natural pesticides. *J. Pest Sci.* **86**: 361-386.
- Biswas, G.C. 2013. Comparative effectiveness of neem extracts and Synthetic organic insecticide against mustard aphid. *Bangladesh J. Agril. Res.* **38(2)**: 181-187.
- Blackman, R. & Eastop, V. 2000. Aphids on the world's crops-an identification and information guide. John Wiley and Sons, U.K.
- Blackman, R. L. & Eastop, V. F. 2007. Biochemical characterization of detoxifying enzymes in Dimethoate-Resistant strains of melon Aphid, *Aphis gossypii* (Hemiptera: Aphidae). *Advances in Entomology.* **4(3)**: 1-29.
- Capinera, J. L. 2001. Handbook of vegetable pests. San Diego, Academic Press. 729 pp.
- Chakraborty, D.P. & Basak S.P. 1971. Cyclomahogenol, a new tetracyclic triterpene from *Swietenia mahagoni*. *Phytochem.* **10(6)**: 1367-1372.
- Cheng, S., Chang, H., Chang, S., Tsai, K. & Chen, W. 2003. Bioactivity of selected plant essential oils against the yellow fever mosquito *Aedes aegypti* larvae. *Bioresource Technol.* **89(1)**: 99-102.
- Cook, B.G., Pengelly, B.C., Brown, S.D., Donnelly, J.L., Eagles, D.A., Franco, M.A., Hanson, J., Mullen, B.F., Partridge, I.J., Peters, M. & Schultze-Kraft, R. 2005. Differential aphid colony establishment in *Dolichos lablab* varieties correlated with some plant specific factors that impact on aphid fecundity. *American J. Plant Sci.* **8(4)**: 754-769.
- Emden, H.F.V. & Harrington, R. 2007. Aphids as crop pests. Oxford, UK, CAB International.
- Franck, E.D., Charles, L.C. & Stephen, O.D. 2009. Natural products in crop protection. *Bioorg. Medic. Chem.* **17(22)**: 4022-4034.
- Gahukar R. 2000. Use of neem products/pesticides in cotton pest management. *Int. J. Pest Manage.* **46**: 149-160.
- Gorur, G., Abdullah, M.I. & loik, M. 2008. Insecticidal activity of the *Thymus*, *Veronica* and agrimonia's essential oils against the cabbage aphid, *Brevicoryne brassicae*. *Acta Phytopathologica et Entomologica Hungarica.* **43(1)**: 201-208.
- Hajra, S., Mehta, A. & Pandey, P. 2011. Phenolic compounds and antioxidant activity of *Swietenia mahagoni* seeds. *Int. J. Pharm. Sci.* **3(Suppl 5)**: 431-434.
- Hold, K.M., Sirisoma, N.S., Ikeda, T., Narahashi, T. & Casida J.E. 2000. Thujone (the bioactive component of absinthe): Aminobutyric acid type a receptor modulation and metabolic detoxification. *Proc. Nat. Academy Sci, USA* **97**: 3826-3831.
- Hossain, M.A., Alim, M.A. Ahmed, K.S. & Haque, M.A. 2014. Insecticidal potentials of plant oils against *Callosobruchus chinensis* (Coleoptera: Bruchidae) in stored chickpea. *J. Entomol. Soc. Iran.* **34(3)**: 47-56.
- Iqbal, M.F., Kahloon, M.H., Nawaz, M.R. & Javaid, M.I. 2011. Effectiveness of some botanical extracts on wheat aphids. *J. Animal & Plant Sci.* **21(1)**: 114-115
- Isman M.B. 2000. Plant essential oils for pest and disease management. *Crop Prot.* **19**: 603-608.
- Isman, M.B. 2006. Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world. *Ann. Rev. Entomol.* **51**: 45-66.
- Jastad, G., Trandem, N., Hovland, B. & Mogan, S. 2009. Effect of botanically derived pesticides mired pest and beneficial in apple. *Crop Pro.* **12**:141-147.
- Kala, K., Sories, T., Mohan, V.R., & Vaduvel, V. 2010. Nutrient and chemical composition of raw seeds of five varieties of *Lablab purpureus* (L.) *Sweetness Adv. Biores.* **1**: 44-53.
- Kamphuis, L.G., Gao, L. & Singh, K.B. 2012. Identification and characterization of resistance to cowpea aphid (*Aphis craccivora* Koch) in *Medicago truncatula*. *BMC Plant Biol.* **12 (101)**: 1-12.
- Karungi, J., Adipala, E., Ogenga-Latigo, M.W.,

- Kyamanywa, S. & Oyobo, N. 2000. Pest management in cowpea. Part 1. Influence of planting time and plant density on cowpea field pest infestation in Eastern Uganda. *Crop Pro.* **19** (4): 231-236.
- Katsvangwa, C.A.T. & Chigwaza, S. 2004. Effectiveness of natural herbs, fever tea (*Lippia javanica*) and Mexican marigold (*Tagetes minuta*) as substitutes to synthetic pesticides in controlling aphid species *Brevicoryne brassicaea* on cabbage (*Brassica capitata*). *Trop. Sub. Trop. Agroecosys.* **4**:101-106.
- Kedia, A., Prakash, B., Mishra, P.K., Singh, P. & Dubey, N.K. 2015. Botanicals as eco-friendly biorational alternatives of synthetic pesticides against *Callosobruchus* spp. (Coleoptera: Bruchidae)- a review. *J. Food Sci. Technol.* **51**(9): 2210-2215.
- Ketoh G.K. 2004. Evaluation of three plants potentialities in field and house termites control in Togo. *Int. Foundation Sci. (IFS) Report.* 27p.
- Khater, H.F. 2012. Prospects of botanical biopesticides in insect pest management. *Pharmacologia.* **3**(12): 641-656.
- Klingler, J., Kovalski, I., Silberstein L., Thompson G. & Perl-Treves, R. 2001. Mapping of cotton-melon aphid resistance in melon. *J. American Soc. Hort. Sci.* **126**(1): 56-63.
- Lee, P. & Tai, D.Y.H. 2001. Clinical features of patients with acute organophosphate poisoning requiring intensive care. *Intensive Care Medicinal.* **27**(4): 694-699.
- Lin, C.Y., Wu, D.C., Yu, Z.H., Chen, B.H. Wang, C.L. & Ko, W.H. 2009. Control of silverleaf whitefly, cotton aphid and kanzawa spider mite with oil and extracts from seeds of sugar apple. *Neotrop. entomol.* **38**(4): 553-536.
- Lowery, D. & Isman, M. 1995. Toxicity of neem to natural enemies of aphids. *Phytoparasitica.* **23**: 297-306.
- Madahi, K., Sahragard, A. & Hossieni, R. 2013. Prey density dependent life table of *aphidoletes aphidomyza* rondani (Diptera: Cecidomyiidae) feeding on *Aphis craccivora* Koch (Hemiptera: Aphididae) under laboratory conditions. *J. Plant Prot. Res.* **53**(3): 253-262.
- Mahmud, M.K., Khan, M.M.H., Husain, M., Alam, M.I. & Afrad, M.S.I. 2002. Toxic effects of different plant oils on pulse beetle *Callosobruchus chinensis* Linn. (Coleoptera: Bruchidae). *J. Asiat. Soc. Bangladesh.* **28**(1): 11-18.
- Manzoor, M., Ali, H., Khalid, S.H., Idrees, A. & Arif, M. 2015. Potential of Moringa (*Moringa oleifera*: Moringaceae) as plant growth regulator and bio-pesticide against wheat aphids on wheat crop (*Triticum aestivum*; Poaceae). *J. Biopest.* **8**: 120-127.
- McDonald, L.L., Guy, R.H. & Speirs, R.D. 1970. Preliminary evaluation of new candidate materials as toxicants repellents and attractants against stored product insects. Marketing Research Report Number 882. Washington.
- Megersa, A. 2016. Botanicals extracts for control of pea aphid (*Acyrtosiphon pisum*; Harris). *J. Entomol. Zool. Stud.* **4**(1): 623-627.
- Oliveira, C.M., Auad, A.M., Mendes, S.M. & Frizzas, M.R. 2014. Crop losses and the economic impacts of insect pests on Brazilian agriculture. *Crop Protect.* **56**: 50-54.
- Patil, D.S. & Chavan, N.S. 2009. Bioefficacy of some botanicals against the sugarcane woolly aphid, *Ceratovacuna lanigera* Zehnter. *J. Biopestic.* **2**(1): 44-47.
- Pavela, R., Nadezda, V. & Bozena, S. 2009. Repellency and toxicity of three Impatiens species (Balsaminaceae) extracts on *Myzus persicae* Sulzer (Homoptera: Aphididae). *J. Biopestic.* **2**(1): 48-51.
- Peris, N.W. & Kiptoo, J.J. 2017. Potential of botanical extracts in the control of kale aphids (*Brevicoryne brassicaea*) and their effect on the parasitic wasp (*Aphidius ervi*) Asian Res. *J. Agricult.* **4**(3): 1-6.
- Pinto, E.S., Barros, E.M., Torres, J.B. & Neves, R.C.S. 2013. The control and protection of cotton plants using natural insecticides against the colonization by *Aphis gossypii* Glover (Hemiptera: Aphididae). *Acta Scientiarum.* **35**(2): 169-174.

- Prakash A. & Rao J. 1996. Botanical Pesticides in Agriculture, CRC Press, New Delhi, India, 480p.
- Rajappan, K., Ushamalini, C., Subramamian, N., Narasimhan, V. & Kareem, A. 2000. Effect of botanicals on the population dynamics of *Nephotettix virescens*, rice tungro disease incidence and yield of rice. *Phytoparasitica*. **28(2)**: 109-113.
- Rawleigh, S.O. & Boyd, A.E. 2008. Compression of Home Made and Conventional Sprays as Aphid Control on Lettuce. *J. North Carolina Acad. Sci.* **124(2)**: 53-57.
- Razaq, M., Mehmood, A., Aslam, M., Ismail, M., Afzal, M. & Alishad, S. 2011. Losses in yield and yield components caused by aphids to late sown *Brassica napus*, *Brassica juncea* and *Brassica carinata* braun at multan, Punjab (Pakistan). *Pakistan J Bot.* **43(1)**: 319-324.
- Sahgal, G., Ramanathan, S., Sasidharan, S., Mordi, M.N., Ismail, S. & Mansor, S.M. 2009. Phytochemical and antimicrobial activity of *Swietenia mahagoni* crude methanolic seed extract. *Trop. Biomed.* **26(3)**: 274-279.
- Saikia, P., Das, D. & Saikia, L. 2000. Evaluation of botanicals and fish oil formulation against bean aphid, *Aphis craccivora* Koch. *J. Agric. Sci. Soc.* **13**: 79-80.
- Sainsbury, F., Cañizares, M.C. & Lomonosoff, G.P. 2010. Cowpea mosaic virus: the plant virus-based biotechnology workhorse. *Ann. Rev. Phytopathol.* **48**: 437-455.
- Salim, M., Hossain, S., Alam, S., Rashid, J.A. & Islam, S. 2013. Estimation of genetic divergence in lablab bean (*Lablab purpureus* L.) genotypes. *Bangladesh J. Agric. Res.* **38 (1)**: 105-114.
- Saranya, S., Ushakumari, R., Jacob, S. & Philip, B.M. 2010. Efficacy of different entomopathogenic fungi against cowpea aphid, *Aphis craccivora* (Koch). *J. Biopestic.* **3 (1)**: 138-142.
- Schoonhoven, A.V. 1978. Use of vegetable oils to protect stored beans from bruchid attack. *J. Econ. Entmol.* **71**: 254-256.
- Shaahu, D.T., Kaankuka, F.G. & Okpanachi, U. 2015. Proximate, amino acid, anti-nutritional factor and mineral composition of different varieties of raw *lablab purpureus* seeds. *Int. J. Sci. Technol. Res.* **4**: 157-161.
- Shah, F.M., Razaq, M., Ali, A., Han, P. & Chen, J. 2017. Comparative role of neem seed extract, moringa leaf extract and imidacloprid in the management of wheat aphids in relation to yield losses in Pakistan. *PLoS ONE.* **12(9)**: 1-24.
- Singh, G., Singh, N.P. & Singh, R. 2014. Food plants of a major agricultural pest *Aphis gossypii* Glover (homoptera: aphididae) from India: an updated checklist. *Int. J. Life Sci. Biotechnol. Pharmaceut. Res.* **3(2)**: 1-26.
- Smith, C.M. & Boyko, E.V. 2007. The molecular bases of plant resistance and defense responses to aphid feeding: current status. *Entomol. Experiment. t Applic.* **122(1)**: 1-16.
- Stapel, J.O., Cortesero, A.M. & Lew, W.J. 2000. Disruptive sub lethal effects of insecticides on biological control: Altered foraging ability and life span of a parasitoid after feeding on extra floral nectar of cotton treated with systemic insecticides. *Bio Control.* **17**: 243-249.
- Suthisut, D., Fields, P.G. & Chandrapatya, A. 2011. Fumigation toxicity of essential oils from three Thai plants (Zingiberaceae) and their major compounds against *Sitophilus zeamais* and *Tribolium castaneum*. *J. Stored Prod. Res.* **47(3)**: 222-230.
- Talukder, F.A. & Howse, P.E. 1995. Evaluation of *Aphanamixis polystachya* as a source of repellents, antifeedants, toxicants and protectants in storage against *Tribolium castaneum* (Herbst). *J. Stored Prod. Res.* **31(1)**: 55-61.
- Thejaswi, L., Mohan, I., Naik, S.N. & Manjunatha, M. 2008. Studies on population dynamics of pest complex of field bean (*Lablab purpureus* L.) and natural enemies of pod borers. *Karnataka J. Agric. Sci.* **21(3)**: 399-402.